PHYTOREMEDIATION OF OXIDATIVE STRESS BY ROOT EXTRACT OF ACHYRANTHES ASPERA IN LYMPHOCYTES OF ORYCTOLAGUS CUNICULUS L.

NISREEN HUSAIN^{a1} AND ANIL KUMAR^b

^aDepartment of Zoology, Govt. Dr. W.W. Patankar Girls' PG. College, Durg, Chhattisgarh, India ^bDepartment of Zoology / Biotechnology, Govt. V.Y.T.PG. Autonomous College, Durg, Chhattisgarh, India

ABSTRACT

Oxidative stress plays an important role in the various pathological processes leading to many dreadful diseases. It arises due to the excessive damages of bio membranes and macromolecules caused by the harmful effects of free radicals. Free radicals (ROS & RNS) are generated during cellular oxidation and metabolic processes. Their decomposition exhibits deleterious effects on various physiological systems of living organisms and human health. The free radicals are scavenged by the cellular antioxidants that constitute the strong antioxidant defense system of the body. The antioxidants from natural sources such as the medicinal plants have been of prime importance in the biological world. The aim of the present study is to evaluate the antioxidant activity of the root extract of Achyranthes aspera, a well-known medicinal herb, for the phyto- remediation of hydrogen peroxide induced oxidative stress in lymphocytes of Oryctolagus cuniculus L. (rabbit). Incubation with H₂O₂ (hydrogen peroxide) for 2 hours increased lipid peroxidation, affecting the concentrations of antioxidant enzymes and glutathione, in terms of malondialdehyde (MDA – 3.90 \pm 0.04), superoxide dismutase (SOD – 1.36 \pm 0.04), reduced glutathione (GSH – 2.36 \pm 0.06), catalase $(CAT - 3.73 \pm 0.02)$ and glutathione peroxidase $(GPx - 4.83 \pm 0.09)$. Pretreatment with the methanolic root extract of Achyranthes aspera for 18 hours was found to control lipid peroxidation and positively enhances the activities of antioxidant enzymes, with gradual increase in dosages, ultimate being 20μ / 10,000 cells, viz., MDA (1.00 ± 0.03), GSH (5.12 ± 0.12), SOD (1.75 ± 0.02), CAT (5.03 ± 0.05) and GPx (8.45 ± 0.36). The high antioxidant activity and the efficient free radical scavenging ability of the methanolic root extract of Achyranthes aspera contributed to the minimization of oxidative stress in lymphocytes. The rich phytochemical content present in the roots is usually considered to be responsible for the antioxidant efficiency of root extract of Achyranthes aspera, and hence ideal for phytoremediation of oxidative stress.

KEYWORDS: Lipid peroxidation, Oxidative stress, Antioxidants, Free radicals

The cellular oxidation and free radical generation leads to impaired antioxidant defense system. The oxygen free radicals induce damage to biomembranes and biomolecules such as proteins, lipids, carbohydrates and DNA (Young et al., 2001 ; Mc Cord et al., 2000), which are responsible for various kinds of cellular dysfunctions. The imbalance and the deleterious effects developed due to free radicals, in the living system, give rise "Oxidative Stress" (Kovacic et al., 2001; to Ridnour et al., 2005). The continuous and rapid production of free radicals leads to deficient antioxidants. Once the natural antioxidant defense system becomes weak, there is biochemical lesion in many metabolic pathways (Sood et al., 2004). Thus, the conditions of oxidative damage and oxidative stress are responsible for pathological processes of various dreadful diseases and fast ageing as well (Tiwari et al., 2001; Mohammad et al., 2004).

The strong and efficient antioxidant system is significantly required for the control of the toxic and harmful state caused by oxidative stress. Antioxidants are the substances that scavenge the free radicals, and thus reduce oxidative stress by inhibiting cellular damages and lipid peroxidation (Irshad *et al.*, 2002), thereby preventing the condition of pathogenecity (Rock *et al.*, 1996). The harm caused to significant antioxidant enzymes of the antioxidant system, *viz.*, MDA (Malondialdehyde), SOD (Superoxide dismutase), CAT (Catalase), GSH (Reduced Glutathione) and GPx (Glutathione peroxidase) is compensated by the routine intake of antioxidants through food, tonics, supplements and medicines.

There are many plants with medicinal values that have been the major part of the traditional medicines since ages, as well as form the basis of many of the modern pharmaceuticals. The medicinal value of such plants has been attributed to their rich phytochemical constituents and antioxidant properties. Thus in recent era, there is an inclination to use antioxidants from natural sources such as medicinal and dietary plants (Brown *et al.*, 1998; Herrera *et al.*, 2001) and their characterization for successful phytoremediation of oxidative stress.

In the string of such antioxidant sources, one of the very commonly available plants in India is *Achyranthes aspera*. It has been traditionally used in the treatment of many cutaneous, respiratory and digestive disorders (Nadkarni, 2009; Pankaj, 2003). The rich antioxidant activity and medicinal property of the root extract of *Achyranthes aspera* is reported due to the presence of phytochemical compounds such as saponin, sterols, alkaloids and polysaccharides (Priya *et al.*, 2010).

In human population, the lymphocytes are considered to be important for encountering the various toxicants and free radicals. The lymphocytes are used in the present study for inducing H_2O_2 into them as the hydroxyl radical formation is easily stimulated in them, leading to oxidative stress. The present paper focuses on the evaluation of antioxidant efficacy of the methanolic root extract of *Achyranthes aspera* to assess phytoremediation of H_2O_2 induced oxidative stress in lymphocytes of *Oryctolagus cuniculus*.

MATERIALS AND METHODS

The healthy plants of *Achyranthes aspera* were collected from the open fields and grounds of Durg (Chhattisgarh), and the roots were separated. After proper washing and sterilization in 70% alcohol, the roots were shade dried. The extract was prepared in 59% methanol by Soxhlet extraction apparatus.

For *in vitro* study, blood sample was collected from *Oryctolagus cuniculus*, and stored in heparinized sterilized tube. This was followed by centrifugation for isolation of lymphocytes and thereafter washing in phosphate buffer saline.

	Group I (Control)	Group II (H ₂ O ₂ treated)	Group III (5µl ARE +	Group IV (10µl ARE +	Group V (20µl ARE +
		$(\Pi_2 O_2 \text{ treated})$	(5µ1 AKE + H ₂ O ₂)	H_2O_2	(20µ1 AKE + H ₂ O ₂)
Lipid peroxides in mole MDA/mg protein (MDA)	0.80 ± 0.02	3.90 ± 0.04*	3.02 ± 0.06*	2.19 ± 0.015#	1.00 ± 0.03#
Reduced Glutathione (µ moles / mg protein) (GSH)	5.96 ± 0.04	2.36± 0.06*	2.72 ± 0.05#	3.37 ± 0.13#	5.12 ± 0.12#
Superoxide dismutase (units / mg protein) (SOD)	3.88 ± 0.06	1.36 ± 0.04*	2.68 ± 0.11#	2.70 ± 0.14#	2.75 ± 0.12#
Catalase (µ moles of H ₂ O ₂ consumed / min./ mg protein) (CAT)	5.56 ± 0.10	3.73 ± 0.02*	3.88 ± 0.03#	4.09 ± 0.04#	5.03 ± 0.05#
Glutathione peroxidase (µg of Glutathione utilized / min. / mg protein) (GPx)	9.56 ± 0.11	4.83 ± 0.09*	4.86±0.03#	5.61 ± 0.18#	8.45 ± 0.36#

 Table 'A' showing effect of Methanolic Root Extract of Achyranthes aspera on the activity of the antioxidant enzymes and in recovery of Oxidative Stress

DMEM medium alongwith 10% fetal serum was used for the culture of lymphocytes. The

culture was maintained in a humidified CO_2 incubator at 37°C temperature and 5% CO_2 for 18 hours. After incubation, the cells/lymphocytes were exposed to oxidative stress by 100μ M H₂O₂ induced lipid peroxidation for 2 hours (Sohi *et al.*, 2003). The experiment was designed with five groups of cultured lymphocytes for the analysis of each of the considered antioxidant enzymes. The samples taken were in replicates of five. Pretreatment with methanolic root extract of

Achyranthes aspera in gradual increasing dosages as 5μ l, 10μ l, and 20μ l/ 10,000 cells successively in the third, fourth and fifth groups of the experiment aided in evaluation of its antioxidant property and ability of phytoremediation of oxidative stress. The enzymatic parameters taken into consideration for antioxidant assay are:

MDA (Malondialdehyde)	determined by following Okhawa et al. method.		
GSH (Reduced Glutathione)	following Misra et al.method		
SOD(Superoxide dismutase)	following Moron et al. method		
CAT (Catalase)	following Bergmeyer et al. method, and		
GPx (Glutathione)	following Rotruck et al. method		

The collected data for all the antioxidant enzymatic parameters were statistically validated by ANOVA. ARE – Achyranthes Root Extract ; * Compared with Control ; # Compared with H_2O_2 treated Group

RESULTS AND DISCUSSION

The H₂O₂ induced lymphocytes give rise to oxidative stress and the hydroxyl radicals increased the lipid peroxidation by their constant attacks on lipid components of the membrane. This increased the levels of MDA, the final product of peroxide process. The state of oxidative stress so developed decreased the antioxidant activities of GSH, GPx, SOD and CAT. The pretreatment with the methanolic root extract of Achyranthes aspera in gradual and successive increasing dosages (5µl, 10 μ l and 20 μ l/10,000 cells), regulated the activity of MDA, and the antioxidant activity of glutathione as well as other antioxidant enzymes, increased and was restored accordingly. This indicated the state of recovery of lymphocytes from oxidative stress (P<0.05) [Table A].

The fact that Achyranthes aspera is known for its rich medicinal values makes it significant for its use in traditional medicinal system. The plant has been traditionally used in the treatment of asthama, cough, digestive and skin diseases. The paste of roots in water is reported to be effective in curing diarrhoea and eye disorders. The roots of Achyranthes aspera are used by the natives of Chhattisgarh in the form of Herbal Mala and Tabiz in order to treat fever and hasten the process of delivery. This is attributed to its wound healing property. Freshly harvested roots are used in preparation of herbal oil (Pankaj, 2003; Neeru et al., 2006). Infusion of root is known as mild astringent in curing bowel complaints, and juice of root in the treatment of diabetes and mild type of leprosy, bleeding in delivery and menstrual disorders (Chopra *et al.*, 1958 ; 1986; Ojha, 1966).

The whole plant of Achyranthes aspera exhibit good antioxidant effect by presenting the formation of free radicals. Rich source of phytochemicals present in the plant is responsible for its antioxidant properties (Edwin et al., 2008). Ecdysterone and ecdysone isolated from the Methanolic extract of roots and whole plant attributes to its antioxidant potentials (Banerji et al., 1970 ; Ikan et al., 1971 ; Rastogi et al., 2004). A new aliphatic acid responsible for antioxidant properties was isolated from the ethanolic extract of roots (Sharma et al., 2009). Ethanolic extract of root and aerial parts of Achyranthes aspera are also well known for its antibacterial activity (Kumar et al., 2003). Also the alcoholic root extract has been reported to exhibit anti-inflammatory activity (Kumar et al., 2009; Mehta et al., 2009).

Antioxidant activity of leaves and roots have been reported by Gayathri et al. (2009). Malarvili et al. (2009) reported antioxidant efficiency in the seeds of Achyranthes aspera for phytoremediation. Free radical scavenging activity of the ethanolic and aqueous extract of roots were assessed using two methods, viz., DPPH radical scavenging activity and superoxide scavenging activity (Paul et al., 2010). Antioxidant ability of Achyranthes aspera was established by using different extracts of root, stem, leaf and inflorescence, evaluated by DPPH assay. With the increase of time and concentration, evaluation of antioxidant ability was reported to be in order of root > stem > inflorescence > leaf (Beaulah et al., 2011).

Root did not show antioxidant activity in chloroform extract, whereas stem extract showed high radical scavenging potential. In ethyl acetate extract, the antioxidant activity of root was higher than stem. Antioxidant potential of stem and root extracts for phytoremediation was evaluated in methanol and water by measuring DPPH radical scavenging activity. Methanol extract showed high antioxidant activity than that of aqueous extract (Priya *et al.*, 2010).

DPPH and superoxide assay methods explain the antioxidant properties of the plant extracts, but significant direct free radical scavenging enzymes are MDA, GSH, SOD, CAT and GPx. The present work holds its novelty in getting the antioxidant property evaluated comprehensively, considering all directly acting enzymes for free radicals in methanolic root extract of *Achyranthes aspera*. Thus, it was observed that the amleorative effect of antioxidant enzymes was very effective on exposure of methanolic extract of root of *Achyranthes aspera*. Hence, the root extract was found suitable source of phytoremediation of oxidative stress in lymphocytes.

REFERENCES

- Banerji A, Chadha MS. (1970). Insect moulting hormone from Achyranthes aspera. Phytochemistry, 9(7):1691.
- Beaulah G, Mohammed Sadiq A, Jaya Santhi R. (2011). Antioxidant and Antibacterial activity of Achyranthes aspera. Annals of Biological Research, 12(5): 662-670.
- Bergmeyer HV, Gowehu K and Grassel M. (1974). Methods of Enzyme Analysis. *Acad Press New York*, P. 438.
- Brown JE, Rice Evan CA. (1998). Luteolin rich Artichoke extract protects low density lipoprotein from oxidation *in vitro Free Radic Res*, 29 : 247-255.
- Charles Lekhya Priya, Gaurav Kumar, loganathan Karthik, Kokati Venkata Bhaskara Rao. (2010). Antioxidant activity of *Achyranthes aspera* Linn. Stem extracts. *Pharmacologyonline*, 2 : 228-237.
- Chopra RN, Chopra IC, Handa KL, Kapoor LD, Chopra'. (1958) .Indigenous Drug of India, U N Dhar & Sons Pvt. Ltd. Calcutta, 2nd Ed., 493.

- Chopra RN, Nayar SL and Chopra IC. (1986). Glossary of Indian Medical Plants. Council of Scientific and Industrial Research, New Delhi, 185-25.
- Edwin S, Edwin Jarald E, Deb L, Jain A, Kinger H, Dutt KR, Amal Raj A.(2008). Wound healing and Antioxidant Activity of *Achyranthes aspera. Pharmaceutical Biology*, 46:12, 824-828.
- Gayathri DS, Archanah A, Abiramasundari P, Priya V, Uma K, Abirami T.(2009). *Indian Journal of Nutrition and Dietetics*, 46(12): 485-490.
- Herrera E, Barbas C. (2001). Vitamin E : Action, metabolism and perspectives. *J Physiol Biochem*, 57 : 43-56.
- Ikan R, Ravid U, Trosset D, and Shulman E. (1971). Ecdysterone, an insect moulting hormone from Achyranthes aspera (Amaranthaceous). Experientia, 27(5): 504–505.
- Irshad M and Chaudhuri PS. (2002). Oxidant and antioxidant system : Role and significance in human body. *Indian J Exp Biol*, 40 : 1233-1239.
- Kovacic, P., & Jacintho, J.D. (2001). Mechanisms of carcinogenesis; Focus on oxidative stress and electron transfer. Curr. Med. Chem., 8,773-796.
- Kumar H, Singh D, Kushwala SKS, Gupta AK. (2009), Comparison of leaf and root extract of *Achyranthes aspera* for its analgesic activity *Der Pharmacia Lettre*, 1(2): 193-198.
- Malarvili T, Gomathi N. (2009). Effect of *Achyranthes aspera* (Linn) seeds on redox and oxidative status in plasma and selected tissues of rats fed with high doses of fructose, *Biosciences and Biotechnology Research Asia*, 6(2) : 659-664.
- Mc Cord JM. (2000). The evolution of free radicals and oxidative stress. *Am J Med*, 108 : 652-9.
- Mehta FA, Patel BG, Pandya SS, Ahir KB, Patel SB. (2009). *Pharmacologyonline*, 3 : 978-985.

- Misra HP and Fridovich I. (1972). The role of superoxide anion in the autoxidation of epinephrine and a simple assay for superoxide dismutase. J Biol Chem 247(10): 3170-3175.
- Mohammed AA, Ibrahim AA. (2004).Pathalogical roles of reactive oxygen species and their defence mechanism. *Saudi Pharm J* ; 12 : 1-18.
- Moron MS, Depierre JW and Mannervik B. (1979). Levels of glutathione, glutathione redcutase and glutathione reductase and glutathione s-transferase activities in rat lung and liver. *Biochem Biophys Acta*, 582 : 67-68.
- Nadkarni KM, (2009). Indian Materia Medica Bombay Popular Prakashan. Vol I : 21.
- Neeru V, Sharma SK (2006). Post-coital antifertility activity of *Achyranthes aspera* Linn. root. *J Ethnopharmacol*, 107 : 179-181.
- Ojha D, Tripathi SN, Singh G. (1966). Role of an indigenous drug (*Achyranthes aspera*) in the management of reactions in Leprosy : Preliminary observations. *Lepr. Rev.* 37 : 115 120.
- Okhawa H, Ohishi N and Yogi K. (1979). Assay for lipid peroxidation in animal tissue by thiobarbituric acid reaction. *Analyt Biochem*, 95 : 351-354.
- Paul D, De D, Ali KM, Chatterjee K, Nandi DK, Ghosh D. (2010). Contraception. 81(4) : 355-361.
- Pankaj Oudhia. (2001, 2002, 2003). Chirchita or Onga (*Achyranthes aspera* var. perphyristachya Hook F) *Res Note*.
- Ram P. Rastogi, Mehrotra BM. (2004). Compendium of Indian Medicinal plants. Central Drug Research Institute, Lucknow and National Institute of Science Communication and Information Resources, New Delhi. Vol – II, 8.

- Ridnour LA, Isenberg JS, Espey MG, Thomas DD, Roberts DD and Wink DA. (2005). Nitric oxide regulates angeogenesis through a functional switch involving thrombo-spondin-1. *Proc Natl Acad Sci*, USA, 102 : 13147-13152.
- Rock CL, Jacob RA, Bowen PE. (1996). Update of biological characteristics of the antioxidant micronutrients – Vitamin C, Vitamin E and the carotenoids. J Am Diet Assoc, 96 : 693-702.
- Rotruck JT, Pope AL, Ganther H, Swanson AB, Hafeman DG and Hocksira WG. (1973). Selenium : biochemical role as a component of glutathione peroxidase. *Science*, 179 : 588-590.
- Sharma P, Bhat T. (2009). DPPH antioxidant assay revisited. *Food Chem*, 113(4) : 1202-5.
- Sohi KK, Mittal N, Hundal MK and Khanduja KI. (2003). Gallic acid, an antioxidant, exhibits antiapoptotic potential in normal human lymphocytes : A B Cl-2 independent mechanism. J Nutr Sci Vitaminol, 49(4) : 221-227.
- Sood PP, Vijaylaxmi K, Bapu C, Chundawat RS and Tyagi S. (2004). Myelin degeneration viz. a viz. inhibition of myelin marker enzyme, CNPase, in the central nervous system – methyl mercury intoxicated mice and protection during therapy. J Cell Tissue Res. Vol – 4(1): 71-74.
- Tiwari AK. (2001). Imbalance in antioxidant defence and human diseases : Multiple approach of natural antioxidant therapy. *Curr Sci*, 81(9) : 1179-1187.
- Young IS, Woodside JV. (2001). Antioxidants in health and disease. *J Clin Pathol*, 54 : 176-86.